

# Package ‘fossilbrush’

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**Title** Automated Cleaning of Fossil Occurrence Data

**Version** 1.0.5

**Description**

Functions to automate the detection and resolution of taxonomic and stratigraphic errors in fossil occurrence datasets. Functions were developed using data from the Paleobiology Database.

**License** GPL (>= 3)

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add_itp	<i>add_itp</i>
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---

### Description

Function to add detected peaks using the output of

### Usage

```
add_itp(x, taxon, legend.pos = "topright", exit = TRUE)
```

**Arguments**

x	The list output of @seealso threshold_ranges
taxon	A character vector of length one, specifying one of the taxon names in x to be plotted
legend.pos	One of topleft, bottomleft, topright or bottomright, or a vector of length two, giving the xy coordinates of the legend. A convenience parameter so that the plot detail can remain unobscured.
exit	Restore base plotting parameters on function exit (default as a requirement for CRAN). Can be set to false to allow other elements to be added to a plot

**Value**

None, the detected peaks are added to an existing density plot

**See Also**

threshold\_ranges. This function should be used to add information to an existing plot from @seealso densify, ensuring that the same taxon name is being used

**Examples**

```
# load dataset#
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# densify ranges
dens <- densify(brachios)
# interpeak thresholding
itp <- threshold_ranges(brachios, win = 8, thresh = 10,
                        rank = "genus", srt = "max_ma", end = "min_ma")
# append the stratigraphically thresholded taxon names to the dataset
# plot the taxon, now identifying the peaks
plot_dprofile(dens, "Atrypa", exit = FALSE)
add_itp(itp, "Atrypa")
```

---

 add\_kingdoms

*add\_kingdoms*


---

**Description**

Convenience function to add in a kingdom column to a PBDB dataset. This relies on the dataset having a column of phylum-level assignments for occurrences. The kingdom column is a useful addition for filtering very large taxonomically diverse datasets, and adds in an additional level of data which can inform taxonomic cleaning routines like those called by @seealso check\_taxonomy

**Usage**

```
add_kingdoms(x, phylum = "phylum", insert.left = TRUE)
```

**Arguments**

<code>x</code>	A dataframe containing, minimally, phylum-level assignments of the data
<code>phylum</code>	A character of length 1 specifying the column in <code>x</code> with the phylum level assignments
<code>insert.left</code>	A convenience argument which will make sure that the kingdom column will be inserted in dataframe left immediately to the left of the phylum column

**Value**

The dataframe `x`, with the kingdom column inserted

**Examples**

```
# load dataset
data("brachios")
# add kingdoms to dataset
brachios <- add_kingdoms(brachios)
```

---

age\_ranges

*roxygen documentation*

---

**Description**

age\_ranges

**Usage**

```
age_ranges(
  data,
  taxonomy = "genus",
  srt = "max_ma",
  end = "min_ma",
  mode = "max"
)
```

**Arguments**

<code>data</code>	A three column dataframe comprising one or more character columns of taxonomic names, a numeric column of FADs and a numeric column of LADs
<code>taxonomy</code>	A character vector corresponding to one or more of the taxonomic name columns in data
<code>srt</code>	A character vector of length one specifying the FAD column in data

end                    A character vector of length one specifying the LAD column in data

mode                   A character vector of length one specifying the type of range table to return: one of max, min or bounds. If not specified by the user, the function behaviour will default to max

**Details**

Function to derive a range table of taxon names from a stratigraphic occurrence dataset. The default behaviour is to return a total range table - the oldest FAD and youngest LAD for each taxon (max), but the function can also return the minimum range - youngest FAD and oldest LAD (min), or the uncertainty bounds on each FAD and LAD - the two oldest FADs and two youngest LADs (bounds). The names for which ranges are derived are specified by the taxonomy argument, but multiple elements can be given here, allowing taxonomic range for higher clades to also be returned.

**Value**

A dataframe containing at least four columns: taxon name, FAD, LAD and the taxonomic rank. If taxonomy is of length one, taxonomic rank will be a vector of identical names. If mode = "bounds", there will be two pairs of age columns, denoting the upper and lower bounds on the FAD and LAD for each taxon name

**Examples**

```
# load dataset
data("brachios")
# derive age ranges
rng <- age_ranges(brachios)
```

---

assess\_duplicates            *assess\_duplicates*

---

**Description**

Function to assess and resolve elements with multiple higher classifications in a tgraph object. Assessment is performed based on the topology of the graph they form. Linear paths (i.e. two totally separate paths diverging from the a shared node), rings (divergent paths which only reunite at the highest rank in the tgraph) or more than two divergent paths are treated as distinct. If not any of these cases, the distance between the focal element and the reunion of the divergent paths, along with their subtologies are assessed and a consensus or preferred path based on the frequency of each path in the tgraph or their completeness returned, or the element judged as having multiple distinct classifications

**Usage**

```
assess_duplicates(
  x,
  node,
  mode = c("frequency", "completeness"),
```

```

    jump = 3,
    plot = FALSE
  )

```

### Arguments

x	A tgraph object
node	character vector of elements with multiple higher classifications in x, or a tvert-seq object with those same elements as focal
mode	The rule to be used in selecting between multiple higher classifications. It is possible for the most complete pathway to also be the most frequent
jump	The maximum number of levels between the point of divergence and the point of reunion (if present) for a given path, below which the divergence will be taken as conflicting
plot	A logical specifying if the divergent paths should be plotted

### Value

A list with as many items as elements with multiple classifications, each recording the assessment for a given element

---

brachios

*brachios*


---

### Description

An example dataset of Palaeozoic brachiopods downloaded from the Paleobiology Database.

### Usage

```
brachios
```

### Format

An object of class `data.frame` with 151473 rows and 10 columns.

---

`check_taxonomy``check_taxonomy`

---

## Description

Wrapper functions to implement a multi-step cleaning routine for hierarchically structured taxonomic data. The first part of the routine calls `@seealso format_check` to perform a few presumptive checks on all columns, scanning for non-letter characters and checking the number of words in each string. By default, `@seealso clean_name` is called to ensure correct formatting as this improves downstream checking. The second part of the routine calls `@seealso spell_check` to flag spelling discrepancies between names within a given taxonomic group. If chosen, the function can automatically impose the more frequent spelling. The third part of the routine calls `@seealso discrete_ranks` to flag name re-use at different taxonomic levels. Some of these cases may arise when a name has been unfortunately, (although permissibly) used to refer to groups at different taxonomic levels, or where a higher classification may have been inserted as a placeholder for a missing lower classification. The fourth part of the routine calls `@seealso find_duplicates` to flag variable higher classifications for a given taxon, including cases where a higher classification is missing for one instance of a taxon, but present for the others. If chosen, `@seealso resolve_duplicates` is called to ensure a consistent classification is imposed. For cases where a name has been re-used at the same rank for genuinely different taxa (not permissible, unlike name re-use at different ranks) suffixes are added as capital letters, e.g. TaxonA, TaxonB. If any of the automatic cleaning routines are employed (again the default behaviour as `clean_name` is TRUE by default), the function will return a cleaned version of the dataset. If the use of suffixes from `@seealso resolve_duplicates` is not desirable, the function behaviour can be altered so that any suffixes are dropped before returning.

## Usage

```
check_taxonomy(  
  x,  
  ranks = c("phylum", "class", "order", "family", "genus"),  
  species = FALSE,  
  species_sep = NULL,  
  routine = c("format_check", "spell_check", "discrete_ranks", "find_duplicates"),  
  report = TRUE,  
  verbose = TRUE,  
  clean_name = FALSE,  
  clean_spell = FALSE,  
  thresh = NULL,  
  resolve_duplicates = FALSE,  
  append = TRUE,  
  term_set = NULL,  
  collapse_set = NULL,  
  jw = 0.1,  
  str = 1,  
  str2 = NULL,  
  alternative = "jaccard",  
  q = 1,  
)
```

```

    pref_set = NULL,
    suff_set = NULL,
    exclude_set = NULL,
    jump = 3,
    plot = FALSE
  )

```

## Arguments

x	A dataframe with hierarchically organised taxonomic information. If x only comprises the taxonomic information, @param ranks does not need to be specified, but the columns must be in order of decreasing taxonomic rank
ranks	The column names of the taxonomic data fields in x. These must be provided in order of decreasing taxonomic rank
species	A logical indicating if x contains a species column. As the data must be supplied in hierarchical order, this column will naturally be the last column in x and species-specific spell checks will be performed on this column. NOTE that for the function to work, the species name must be the full species name rather than just the specific epithet, e.g., 'Tyto_alba' rather than just 'alba'.
species_sep	A character vector of length one specifying the genus name and specific epithet in the species column <ul style="list-style-type: none"> <li>• Flagging routine arguments *</li> </ul>
routine	A character vector determining the flagging and cleaning routines to employ. Valid values are format_check (check for non letter characters and the number of words in names), spell_check (flag potential spelling errors), discrete_rank (check that taxonomic names are unique to their rank), duplicate_tax (flag conflicting higher classifications of a given taxon)
report	A logical of length one determining if the flagging outputs of each cleaning routine should be returned to the user for inspection. This is different to @param verbose, which controls whether flagging should additionally be reported to the user on the console
verbose	A logical determining if function progress and flagged errors should be reported to the console <ul style="list-style-type: none"> <li>• Cleaning routine arguments *</li> </ul>
clean_name	If TRUE, the function will return cleaned versions of the columns in x using the routines in @seealso clean_name. These routines can be altered using the 'term_set' and 'collapse_set' arguments.
clean_spell	If TRUE, the function will return a cleaned version of the supplied taxonomic dataframe, using the supplied threshold for the similarity method given by method2, to automatically update any names in pairs of flagged synonyms to the more frequent spelling. This is not recommended, however, so the argument is FALSE by default and the threshold left as NULL
thresh	The threshold for the similarity method given by method2, below which flagged pairs of names will be considered synonyms and resolved automatically. See @seealso spell_check for details on method2



resolve_duplicates	If TRUE, the function will return a cleaned version of the supplied taxonomic dataframe, using @seealso resolve_duplicates to resolve conflicts in the way documented by the function. Both spell_clean and tax_clean can both be TRUE to return a dataset cleaned by both methods
append	If TRUE, any suffixes used during cleaning will be retained in the cleaned version of the data. This is preferable as it ensures that all taxonomic names are rank-discrete and uniquely classified <ul style="list-style-type: none"> <li>• Routine specific arguments *</li> </ul>
term_set	A character vector of terms (to be used at all ranks) or a list of rank-specific terms which will be supplied, element-wise as the @param collapse argument called by @seealso clean_name. If a list, this
collapse_set	A character vector of character strings (to be used at all ranks) or a list of rank-specific strings which will be supplied, element-wise as the @param collapse argument called by @seealso clean_name. If a list, this should be given in descending rank order
jw	Called by @seealso spell_check
str	Called by @seealso spell_check
str2	Called by @seealso spell_check
alternative	Called by @seealso spell_check
q	Called by @seealso spell_check
pref_set	A character vector of prefixes (which will be used at all ranks) or a list of rank-specific prefixes, which will be supplied, element-wise as the @param pref argument called by @seealso spell_check. If a list, this should be given in descending rank order.
suff_set	A character vector of suffixes (which will be used at all ranks) or a list of rank-specific suffixes, which will be supplied, element-wise as the @param suff argument called by @seealso spell_check. If a list, this should be given in descending rank order.
exclude_set	A character vector of terms to exclude (which will be used at all ranks) or a list of rank-specific exclusion terms, which will be supplied, element-wise as the @param exclude argument called by @seealso spell_check. If a list, this should be given in descending rank order.
jump	Called by @seealso resolve_duplicates
plot	Called by @seealso resolve_duplicates

### Details

- Data supply arguments \*

### Value

A list with elements corresponding to the outputs of the chosen flagging routines (four by default: \$formatting, \$synonyms, \$ranks, \$duplicates), plus a cleaned version of the data (\$data) if any of clean\_name, clean\_spell or resolve\_duplicates are TRUE. See @seealso format\_check, @seealso spell\_clean,

**See Also**

discrete\_ranks and @seealso find\_duplicates for details of the structure of the flagging outputs

**Examples**

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# define the taxonomic ranks used in the dataset (re-used elsewhere)
b_ranks <- c("phylum", "class", "order", "family", "genus")
# define a list of suffixes to be used at each taxonomic level when scanning for synonyms
b_suff = list(NULL, NULL, NULL, NULL, c("ina", "ella", "etta"))
# scan for errors
brachios <- check_taxonomy(brachios, suff_set = b_suff, ranks = b_ranks)
```

---

chronoscale

*chronoscale*

---

**Description**

Convenience function to apply user-specified chronostratigraphy to fossil datasets. The function relies on a lookup table generated based on the named intervals in the PBDB in early 2021. First and last interval names in the supplied dataset are matched against this lookup table, by default using 'get("GTS\_2020)", to get GTS\_2020 numeric ages. If the dataset contains intervals which are not present in the lookup table, they will not be matched and the user will be warned. To get around this possibility, the user can also supply the original numeric ages which will be used as default ages if an interval cannot be matched, to ensure that the returned vectors of numeric ages do not contain NAs.

**Usage**

```
chronoscale(
  x,
  tscale = "GTS_2020",
  srt = "early_interval",
  end = "late_interval",
  max_ma = NULL,
  min_ma = NULL,
  verbose = TRUE
)
```

**Arguments**

x A data.frame containing, minimally two columns corresponding respectively to the first and last intervals of the data. Values should only be present in the second column where the minimum age interval for a row is different to the maximum

	age interval. Otherwise the values should be NA and the ages returned will be based on the interval specified in the first column, in line with PBDB formatting.
tscale	A character string specifying one of the inbuilt chronostratigraphic timescales (currently GTS 2020 only) or a data.frame supplied by the user. If the latter, this must contain columns named 'Interval', 'FAD', 'LAD', specifying the interval names to be matched and their lower and upper age in Ma
srt	A character of length 1 specifying the column name of the first interval field in x
end	A character of length 1 specifying the column name of the last interval field in x
max_ma	If not NULL, a character of length 1 specifying the column name of the original numeric maximum age field in x, to be used as fall back values if interval names cannot all be matched
min_ma	If not NULL, a character of length 1 specifying the column name of the original numeric minimum age field in x, to be used as fall back values if interval names cannot all be matched
verbose	A logical indicating if warning messages should be displayed or otherwise

**Value**

The dataframe, x, with two additional columns containing the revised first and last numeric ages of the data, with column names GTS\_FAD and GTS\_LAD respectively

**Examples**

```
# example dataset
data("brachios")
# add GTS_2020 dates
brachios <- chrono_scale(brachios, srt = "early_interval", end = "late_interval",
                        max_ma = "max_ma", min_ma = "min_ma")
```

---

clean\_name *roxygen documentation*

---

**Description**

clean\_name

**Usage**

```
clean_name(x, terms = NULL, collapse = NULL, verbose = FALSE)
```

**Arguments**

x	a vector of names to clean. This will be coerced to class character internally
terms	a character vector of terms to remove from elements of x. Terms are only removed as whole words, rather than if they also happen to occur as strings within elements of x
collapse	a character vector of strings which should collapsed (i.e. replaced by "", rather than the default " "). If one of the collapse terms is a special regex character, it will need to be escaped, e.g. "\\-"
verbose	A logical of length 1 determining if function progress should be reported to the console

**Details**

Function which bundles a series of cleaning routines into a single process. First any words in brackets are removed, followed by a series of user-defined terms if given. Next Roman and Arabic numerical are removed, then abbreviations up to five letters (abbreviations are matched by the following dot e.g ABFS.). By default, characters for removal are replaced by a white space to prevent accidental collapse of strings. However, there may be specific cases where a collapse is required and so terms given in collapse are dealt with next. After collapsing, rogue all rogue punctuation is removed, then isolated lowercase letters, then isolated groups of capitals up to 5 characters long. Finally, white spaces greater than 1 are removed, along with trailing white space, any remaining strings longer than 2 words subsetted to the first word, the first letter of each string capitalised and zero length strings converted to NA

**Value**

a character vector the same length as x. Elements which were reduced to zero characters during cleaning are returned as NA

**Examples**

```
# load dataset
data("brachios")
# clean genus names
gen_clean <- clean_name(brachios$genus)
```

---

densify

*densify*


---

**Description**

Function to create a matrix of occurrence record densities through geological time from an occurrence dataset. Each column represents a taxon. Each row represents a user defined window of time, with the first row starting at the oldest FAD in the dataset and spanning to the youngest LAD step-wise by the user defined window (default of 1 Ma). Occurrence records are densified by generating a vector of time points from occurrence FAD to occurrence LAD (default step of 0.1 Ma), then

tallied in two ways. The first way is a simple histogram count of points-per-window, with the same number of histogram bins as time steps between the overall taxon FAD and LAD. The second way is a kernel density estimate, using a Gaussian kernel with a equally spaced estimates equal to the number of timesteps between the overall taxon FAD and LAD

### Usage

```
densify(
  x,
  rank = "genus",
  srt = "max_ma",
  end = "min_ma",
  step = 1,
  density = 0.1,
  method = c("histogram", "kernel"),
  ...,
  verbose = TRUE
)
```

### Arguments

<code>x</code>	An occurrence dataset
<code>rank</code>	The column name in <code>x</code> containing the taxon names for which densified columns will be generated
<code>srt</code>	A column name in <code>x</code> denoting the occurrence FADs
<code>end</code>	A column name in <code>x</code> denoting the occurrence LADs
<code>step</code>	A positive integer specifying the time window size (i.e. the duration represented by each row in the output matrix)
<code>density</code>	A positive numeric specifying the step size for densifying records. This should ideally be smaller than <code>step</code>
<code>method</code>	The method for quantifying occurrence density. By default both histogram and kernel density will be used
<code>...</code>	additional arguments passed to <code>@seealso density</code>
<code>verbose</code>	A logical determining if function progress should be reported

### Value

A list of two sparse matrices, the first containing the histogram counts, the second the kernel density estimates

### Examples

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
```

```
# densify ranges
dens <- densify(brachios)
```

---

discrete\_ranks      *roxygen documentation*

---

## Description

discrete\_ranks

## Usage

```
discrete_ranks(x, ranks = NULL)
```

## Arguments

x	A dataframe containing hierarchically structured information, for example a table of genus names and their higher taxonomic classifications
ranks	If not NULL, a vector of column names of x, given in rank order. This is useful if x contains columns which are not rank relevant or if columns are not in hierarchical order. If not supplied, the column order in x is used directly and is assumed to be in rank order

## Details

Function for checking whether names in one column of a hierarchically organised dataframe re-occur at other levels. Two checks are performed. The first checks for names in adjacent column, assuming that accidental reuse of names at other levels are most likely to occur at an adjacent rank. The second compares across all columns.

## Value

A list of two lists. The first list contains names which reoccur at adjacent ranks. The second list contains names that reoccur at any rank

## Examples

```
# load dataset
data("brachios")
# define ranks
b_ranks <- c("phylum", "class", "order", "family", "genus")
# run function
flag <- discrete_ranks(brachios, ranks = b_ranks)
```

---

find_duplicates	<i>find_duplicates</i>
-----------------	------------------------

---

**Description**

Function to detect and report elements with multiple higher assignments in a hierarchically structured dataframe

**Usage**

```
find_duplicates(x, ranks = NULL)
```

**Arguments**

x	A hierarchically organised dataframe
ranks	The ranks in the dataframe in which to check for elements with multiple higher classifications. The top rank is ignored by default

**Value**

A dataframe of elements with multiple higher classifications and their ranks

**Examples**

```
# load dataset
data("brachios")
b_ranks <- c("phylum", "class", "order", "family", "genus")
# run function
flag <- find_duplicates(brachios, ranks = b_ranks)
```

---

find_peaks	<i>find_peaks</i>
------------	-------------------

---

**Description**

Function to scan, column-wise, a matrix of per-taxon observation density time series. This can be applied to either the histogram or the kernel density output of `densify`, but the latter is recommended. Peaks are detected as local maxima, then smoothed within a local window and tested to distinguish if they are noise or significant. Strict threshold is that the peak is greater than the mean + sd of the window

**Usage**

```
find_peaks(x, win = 5, verbose = TRUE)
```

**Arguments**

x	A matrix as outputed by densify
win	A positive integer specifying the window length on either side of a peak (i.e. win 5 will give a total window of 11 - -5 indices + peak index + 5 indices)
verbose	A logical determining if function progress should be reported

**Value**

A list of four, the first three positions containing lists of the peak indices for each taxon, under raw, mean + sd and mean detection regimes. The fourth item is a dataframe of counts of peaks per taxon, 1 row per taxon, 1 column per detection regime

**Examples**

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# get density matrix
dens <- densify(brachios)
# run function, using kernel density matrix
pk <- find_peaks(dens$kdensity)
```

---

flag\_ranges

*flag\_ranges*


---

**Description**

Function to compare stratigraphic ranges in x to a set of reference ranges from y. A list of two elements is returned. The first is a dataframe summarising the overall error status, specific error counts FAD and LAD differences, and the 95% density distributions of the FAD and LAD errors for each unique taxon in the column of x denoted by the first element of xcols. If a taxon in x is not present in y, it is assigned the status 000 and its other entries in the returned dataframe will be NA. The second element of the returned list is the error code for every individual element of the column of x denoted by the first element of xcols - this will have the same number of rows as x. If x is a range table rather than an occurrence dataset, then the two list elements will have the same number of rows. Ranges for comparison may be supplied directly in y, or y may be another occurrence dataset, in which case

**Usage**

```
flag_ranges(
  x = NULL,
  y = NULL,
  xcols = c("genus", "max_ma", "min_ma"),
  ycols = NULL,
```



```

    flag.diff = 5,
    verbose = TRUE
  )

```

### Arguments

x	Stratigraphic range data for taxa as a whole or for individual fossil occurrences
y	The same as in x. This is the dataset to which ranges will be compared
xcols	A character vector of length three specifying, in the following order, the taxonomic name, stratigraphic base (FAD) and stratigraphic top (LAD) columns in x.
ycols	An optional character vector of length three for the same column types as in xcols, but for dataset y. This is useful if the column names differ between the datasets
flag.diff	A vector of thresholds, given in millions of years which will be used to flag discrepancies between occurrence FADs and LADs with respect to the reference range. This is a convenience parameter so that occurrences with large discrepancies can be quickly identified. Multiple thresholds can be supplied
verbose	A logical of length one determining if the flagging progress should be reported to the console

### Value

A list of two data.frames, the first recording overall error statistics, the second recording error types for each element of x. In the second data.frame, FAD or LAD differences in excess of the supplied threshold(s) are marked with 1, otherwise 0

### See Also

age\_ranges is called internally to generate the range table for comparison.

### Examples

```

# load the example datasets
data(brachios)
data(sepkoski)
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# update brachios to GTS2020 to match Sepkoski
brachios <- chrono_scale(brachios, srt = "early_interval", end = "late_interval",
                        max_ma = "max_ma", min_ma = "min_ma", verbose = FALSE)
brachios$max_ma <- brachios$newFAD
brachios$min_ma <- brachios$newLAD
# drop occurrences with older LADs than FADs
brachios <- brachios[brachios$max_ma > brachios$min_ma,]
# trim the Sepkoski Compendium to the relevant entries
sepkoski <- sepkoski[which(sepkoski$PHYLUM == "Brachiopoda"),]
# run flag ranges
flg <- flag_ranges(x = brachios, y = sepkoski, ycols = c("GENUS", "RANGE_BASE", "RANGE_TOP"))

```

---

format_check	<i>format_check</i>
--------------	---------------------

---

### Description

Function to perform a series of basic formatting checks geared towards taxonomic name data. The function very simply checks for non letter characters in the taxonomic names, that species-level names contain two words, and genus-level and above names contain one word.

### Usage

```
format_check(x, ranks, species = FALSE, species_sep = " ", verbose = TRUE)
```

### Arguments

x	A dataframe with hierarchically organised, taxonomic information. If x only comprises the taxonomic information,
ranks	does not need to be specified, but the columns must be in order of decreasing taxonomic rank @param ranks The column names of the taxonomic data fields in x. These must be provided in order of decreasing taxonomic rank
species	A logical indicating if x contains a species column. As the data must be supplied in hierarchical order, this column will naturally be the last column in x and species-specific spell checks will be performed on this column.
species_sep	A character vector of length one specifying the genus name and specific epithet in the species column, if present
verbose	A logical determining if any flagged errors should be reported to the console

### Value

A list of two lists. The first list flags the row indexes of columns whose elements contains non-letter characters. The second list flags the row indexes of columns whose elements do not contain the correct numbers of words

### Examples

```
# load dataset
data("brachios")
# define ranks
b_ranks <- c("phylum", "class", "order", "family", "genus")
# run function
flag <- format_check(brachios, ranks = b_ranks)
```

---

geog_lookup	<i>geog_lookup</i>
-------------	--------------------

---

**Description**

lookup table called by 'get\_pbdb'

**Usage**

geog\_lookup

**Format**

An object of class data.frame with 511 rows and 2 columns.

---

GTS_2020	<i>GTS_2020</i>
----------	-----------------

---

**Description**

lookup table called by 'chrono\_scale'

**Usage**

GTS\_2020

**Format**

An object of class data.frame with 1534 rows and 9 columns.

---

GTS_2020_changelog	<i>GTS_2020_changelog</i>
--------------------	---------------------------

---

**Description**

changelog of periodic updates made to the GTS2020 table originally published in this package. The purpose of this changelog is to allow the user to assess how up-to-date the resource is and made any changes themselves if needed. A data.frame with date-wise rows of edits

**Usage**

GTS\_2020\_changelog

**Format**

An object of class data.frame with 1 rows and 2 columns.

---

intersect_ranges	<i>intersect_ranges</i>
------------------	-------------------------

---

## Description

Function to find the maximum intersection between a set of numeric ranges, in this case first and last appearance datums on taxonomic ranges.

## Usage

```
intersect_ranges(x, srt = NULL, end = NULL, verbose = TRUE)
```

## Arguments

x	A numeric data.frame or matrix of ranges. If just two columns are supplied, the first column is assumed to be the srt column
srt	If x contains more than two columns, srt is the name of the range base column - the FAD
end	If x contains more than two columns, end is the name of the range top column - the LAD
verbose	A logical indicating whether the function should report progress to the console

## Value

A matrix with three columns, indicating the intersection (FAD and LAD) and the number of ranges that intersection encompasses

## Examples

```
# plot an example
df <- cbind(c(1.5, 3, 2.1, 1), c(6, 5, 3.7, 10.1))
plot(1:11, ylim = c(0, 5), col = NA)
segments(x0 = c(1.5, 3, 2.1, 1), y0 = 1:4, x1 = c(6, 5, 3.7, 10.1), y1 = 1:4)
abline(v = 3, col = "red", lty = 2)
abline(v = 3.7, col = "red", lty = 2)
# intersect function
intersect_ranges(df)
```

---

pacmacro\_ranges      *pacmacro\_ranges*

---

### Description

Function to apply a modification of Pacman trimming to macrofossil data. The function generates a densified occurrence record using the same methods as `densify` then trim the upper and lower ranges by a user-defined percentage. The full and trimmed ranges are then compared against each other to test if the FAD and the LAD for a taxon form a long tail in its distribution. Multiple tail thresholds can be supplied, but all test to see if the sum of the FAD and LAD which exceeds the trimmed range constitute the threshold proportion of the total range for than taxon, e.g. does the FAD and the LAD outside of the trimmed range comprise a quarter (`tail.flag = 0.25`) of the taxon range?

### Usage

```
pacmacro_ranges(
  x,
  rank = "genus",
  srt = "max_ma",
  end = "min_ma",
  step = 1,
  density = 0.1,
  top = 5,
  bottom = 5,
  tail.flag = 0.35,
  method = c("histogram", "kernel")
)
```

### Arguments

<code>x</code>	A stratigraphic occurrence dataset
<code>rank</code>	The column name in <code>x</code> containing the taxon names for which trimmed ranges will be calculated
<code>srt</code>	A column name in <code>x</code> denoting the occurrence FADs
<code>end</code>	A column name in <code>x</code> denoting the occurrence LADs
<code>step</code>	A positive integer specifying the time window size (i.e. the duration represented by each row in the output matrix)
<code>density</code>	A positive numeric specifying the step size for densifying records. This should ideally be smaller than <code>step</code>
<code>top</code>	The percentage by which the top of the range will be trimmed
<code>bottom</code>	The percentage by which the bottom of the range will be trimmed
<code>tail.flag</code>	a numeric vector of proportions in the range $0 < x < 1$ which will be used to test for long tails
<code>method</code>	The method for quantifying occurrence density. By default both histogram and kernel density will be used

**Value**

If the user specifies a specific method (e.g. `method = "kernel"`), the returned value will be a `data.frame` containing the taxa as row names, the original taxon ranges (FAD, LAD), their ranges as trimmed by the specified value (default FAD95, LAD95), and the tail status (0 = none, 1 = tail) at the user-specified tail proportions. If method is not specified, the result will be a list of 2 `data.frames`, one for each method

**Source**

Pacman procedure modified from <https://rdr.io/github/plannapus/CONOP9companion/src/R/pacman.R>.

**References**

Lazarus et al (2012) Paleobiology

**Examples**

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# run pacmacro
pacm <- pacmacro_ranges(brachios, tail.flag = c(0.3, 0.35, 0.4),
                        rank = "genus", srt = "max_ma", end = "min_ma")
```

---

pbdb\_fields

*geog\_lookup*

---

**Description**

lookup table called by `'get_pbdb'`

**Usage**

```
pbdb_fields
```

**Format**

An object of class `list` of length 31.

---

pbdb\_kingdoms                      *pbdb\_kingdoms*

---

**Description**

lookup table called by 'get\_pbdb'

**Usage**

pbdb\_kingdoms

**Format**

An object of class list of length 3.

---

plot\_dprofile                      *plot\_dprofile*

---

**Description**

Function to plot density profiles of occurrences through time using the output of @seealso densify.

**Usage**

```
plot_dprofile(x, taxon, exit = TRUE)
```

**Arguments**

x	The list output of @seealso densify
taxon	A character vector of length one, specifying one of the taxon names in x to be plotted
exit	Restore base plotting parameters on function exit (default as a requirement for CRAN). Can be set to false to allow other elements to be added to a plot

**Value**

NULL, the plotted density profile

**Examples**

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# densify ranges
dens <- densify(brachios)
plot_dprofile(dens, "Atrypa")
```

---

plot_taxa	<i>plot_taxa</i>
-----------	------------------

---

### Description

Function to plot the parent or child relationships of an element in a hierarchically organised dataframe. Multiple taxa can be plotted simultaneously

### Usage

```
plot_taxa(  
  x,  
  taxon,  
  trunk,  
  ranks,  
  mode = c("parent", "child", "all"),  
  step = NULL  
)
```

### Arguments

x	a dataframe containing hierarchically organised data in columns
taxon	A character vector of element names whose relationships will be plotted (these must be of the same rank)
trunk	A character vector of length one corresponding to the column name in x in which taxa is located
ranks	A character vector corresponding to the column names in x, given in hierarchical order
mode	The direction of the relationships to be plotted
step	A positive integer specifying the neighbourhood of the relationships to plot. Specifying a number greater than the number of ranks will not cause a failure, and will instead plot all relationships in the direction specified in mode

### Value

A plot of the relationships of the specified elements

### Examples

```
# load dataset  
data("brachios")  
# define ranks in dataset  
b_ranks <- c("phylum", "class", "order", "family", "genus")  
# plot taxon  
plot_taxa(brachios, "Atrypa", trunk = "genus", ranks = b_ranks, mode = "parent")
```



---

```
quantile_coef_density_BMS
      quantile_coef_density_BMS
```

---

### Description

Static rip of the `quantile.coef.density` function and relevant internals from the BMS package as the package is archived.

### Usage

```
quantile_coef_density_BMS(
  x,
  probs = seq(0.25, 0.75, 0.25),
  names = TRUE,
  normalize = TRUE,
  ...
)
```

### Arguments

<code>x</code>	a object of class <code>pred.density</code> , <code>coef.density</code> , <code>density</code> , or a list of densities
<code>probs</code>	numeric vector of probabilities with values in range 0 - 1. Elements very close to the boundaries return <code>Inf</code> or <code>-Inf</code>
<code>names</code>	logical; if <code>TRUE</code> , the result has a <code>names</code> attribute, resp. a <code>rownames</code> and <code>colnames</code> attributes. Set to <code>FALSE</code> for speedup with many <code>probs</code>
<code>normalize</code>	logical if <code>TRUE</code> then the values in <code>x\$y</code> are multiplied with a factor such that their integral is equal to one
<code>...</code>	further arguments passed to or from other methods.

### Value

If `x` is of class `density` (or a list with exactly one element), a vector with quantiles. If `x` is a list of densities with more than one element (e.g. as resulting from `pred.density` or `coef.density`), then the output is a matrix of quantiles, with each matrix row corresponding to the respective density.

### Source

static rip from BMS package

---

resolve\_duplicates      *resolve\_duplicates*

---

### Description

Function for identifying and resolving alternative higher assignments in a hierarchically structured dataframe. Columns are checked from the lowest to the highest rank for elements with multiple higher assignments. These assignments are then assessed topologically to determine if they represent inadvertent use of the same name at a given rank for genuinely different entities, or whether the higher classifications are conflicting. In the case of the former, unique character suffixes are applied to each differently classified case (up to 26 currently supported), effectively splitting up the alternatively classified element. In the case of the latter, the alternative classifications are assessed and are either combined, or the more frequently used or the more complete classification scheme is taken (the more frequent pathway can also be the most complete).

### Usage

```
resolve_duplicates(x, ranks = NULL, jump = 4, plot = FALSE, verbose = TRUE)
```

### Arguments

x	A dataframe containing hierarchically structured information, for example a table of genus names and their higher taxonomic classifications
ranks	If not NULL, a vector of column names of x, given in rank order. This is useful if x contains columns which are not rank relevant or if columns are not in hierarchical order. If not supplied, the column order in x is used directly and is assumed to be in rank order
jump	The maximum number of levels between the point of divergence and the point of reunion (if present) for a given path, below which the divergence will be taken as conflicting
plot	A logical specifying if the divergent paths should be plotted
verbose	A logical of length one which determines if the function should report the detection and resolution of elements with multiple higher classifications (if any)

### Value

The dataframe x, with any alternative higher classifications resolved, giving the classification a strict tree structure

### Examples

```
# load dataset
data("brachios")
# define ranks
b_ranks <- c("phylum", "class", "order", "family", "genus")
# run function
res <- resolve_duplicates(brachios, ranks = b_ranks)
```

---

<code>revise_ranges</code>	<code>revise_ranges</code>
----------------------------	----------------------------

---

## Description

Function to generate a consensus age for assemblages of fossil data in *x*, given a table of taxonomic ranges. The need for error-checking is informed by the error codes for the individual fossil occurrences within each collection - if there is no error, then the consensus age is unchanged. If errors are present, then a consensus age for a threshold proportion of taxa is searched for using the overlap of the ranges for those taxa, as given in range table *y*. Taxa whose occurrences lie outside this consensus age are flagged as potential taxonomic errors. If the threshold consensus partially overlaps with the assemblage age, this overlap is returned to present overzealous alteration of the age - otherwise the complete consensus age is returned. If a consensus age cannot be found, the original assemblage age is returned, and each occurrence in the collection flagged as potential taxonomic errors.

## Usage

```
revise_ranges(
  x,
  y,
  assemblage = "collection_no",
  srt = "max_ma",
  end = "min_ma",
  taxon = "genus",
  err = NULL,
  do.flag = FALSE,
  prop = 0.75,
  allow.zero = TRUE,
  verbose = TRUE
)
```

## Arguments

<code>x</code>	Fossil occurrence data grouped into spatiotemporally distinct assemblages
<code>y</code>	A stratigraphic range dataset from which consensus assemblage ages will be derived
<code>assemblage</code>	The column name of the assemblage groups in <i>x</i>
<code>srt</code>	The column name of stratigraphic bases for each element in both <i>x</i> and <i>y</i> - i.e. <i>x</i> and <i>y</i> must have this same name for that column
<code>end</code>	The column name of stratigraphic tops for each element in both <i>x</i> and <i>y</i> - i.e. <i>x</i> and <i>y</i> must have this same name for that column
<code>taxon</code>	The column name denoting the taxon names in both <i>x</i> and <i>y</i> - i.e. <i>x</i> and <i>y</i> must have this same name for that column

err	The column name flagging age errors for occurrences in x. This allows 100\$ valid assemblages to be skipped. Age errors can be derived using @seealso flag_ranges. All error codes must be one of: "000" - unchecked, "R1R" - valid, "0R0" - both FAD and LAD exceeded, "00R" - totally older than range, "R00" - totally younger than range, "01R" - FAD exceeded, "1R0" - LAD exceeded. If not supplied, all assemblages will be checked, even if they are already valid a priori.
do.flag	Rather than supplying error codes, should flag_ranges be called internally to generate error codes for supply to the rest of revise_ranges? As with err, this is useful to prefilter individual occurrences, allowing assemblages contain all valid, all unchecked or a mixture of such error codes to be skipped. This can massively speed up processing time for large datasets.
prop	A numeric, between 0 and 1, denoting the threshold percentage of taxa in the assemblage for which a consensus age must be found
allow.zero	A logical determining if, in the case of a collection LAD being equal to the consensus age FAD (i.e. a pointwise overlap), that pointwise age will be taken as the revised age. The resultant collection age will have no uncertainty as a result, which may be unrealistic. The default behaviour is FALSE, in which case pointwise overlaps will be ignored and the revised age taken instead
verbose	A logical determining if the progress of the redating procedure should be reported

### Value

A list of two dataframes, the first recording the results of the consensus redating procedure for each assemblage in x, the second recording any flags (if any) for each occurrence in x

### Examples

```
# load datasets
data("brachios")
data("sepkoski")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# rename columns in Sepkoski to match brachios
colnames(sepkoski)[4:6] <- c("genus", "max_ma", "min_ma")
# flag and resolve against the Sepkoski Compendium, collection-wise
revrng <- revise_ranges(x = brachios, y = sepkoski, do.flag = TRUE, verbose = TRUE,
                        taxon = "genus", assemblage = "collection_no",
                        srt = "max_ma", end = "min_ma")
# append the revised occurrence ages and error codes to the dataset
brachios$newfad <- revrng$occurrence$FAD
brachios$newlad <- revrng$occurrence$LAD
brachios$errcode <- revrng$occurrence$status
```

---

sepkoski	<i>sepkoski</i>
----------	-----------------

---

**Description**

An example dataset. A port of the Sepkoski Compendium from the chronosphere package, with a few corrections and GTS2020 dating applied

**Usage**

```
sepkoski
```

**Format**

An object of class `data.frame` with 35700 rows and 6 columns.

---

sep_code	<i>sep_code</i>
----------	-----------------

---

**Description**

Lookup table of chronostratigraphic stage abbreviations used in the Sepkoski Compendium, with interval boundaries updated to the GTS2020 standard

**Usage**

```
data(sep_code)
```

**Format**

An object of class `data.frame` with 306 rows and 8 columns.

**Source**

chronosphere (fetch), Sepkoski 2002

---

 spell\_check

*spell\_check*


---

### Description

Function for checking for potential synonyms with alternate spellings. Synonyms are checked for within group using using a Jaro Winkler string distance matrix. Potential synonyms are selected using the `jw` threshold. These can then be further filtered by the number of shared letters at the beginning and end of the a synonym pair, and by prefixes or suffixes which may give erroneously high similarities.

### Usage

```
spell_check(
  x,
  terms = NULL,
  groups = NULL,
  jw = 0.1,
  str = 1,
  str2 = NULL,
  alternative = "jaccard",
  q = 1,
  pref = NULL,
  suff = NULL,
  exclude = NULL,
  verbose = TRUE
)
```

### Arguments

- |                     |  |
|---------------------|--|
| <code>x</code>      | a dataframe containing a column with terms, and a further column denoting the groups within which terms will be checked against one another. If supplying a dataframe with just these columns, terms should be column 1  |
| <code>terms</code>  | a character vector of length 1, specifying the terms column in <code>x</code> . This is required if <code>x</code> contains more than two columns. Alternatively, if <code>x</code> is not provided, terms can be a character vector. If groups are not specified, all elements of terms will be treated as part of the same group |
| <code>groups</code> | a character vector of length 1, specifying the groups column in <code>x</code> . This is required if <code>x</code> contains more than two columns. Alternatively, if terms is supplied as a character vector, groups can also be supplied in the same way to denote their groups  |
| <code>jw</code>     | a numeric greater than 0 and less than 1. This is the distance threshold below which potential synonyms will be considered   |
| <code>str</code>    | A positive integer specifying the number of matching characters at the beginning of synonym pairs. By default 1, i.e. the first letters must match   |

str2	If not NULL, a positive integer specifying the number of matching characters at the end of synonym pairs
alternative	A character string of length one corresponding to one of the methods used by @seealso afin. One of "osa", "lv", "dl", "hamming", "lcs", "qgram", "cosine", "running_cosine", "jaccard", or "soundex".
q	q-gram size. Only used when alternative is "qgram", "cosine" or "Jaccard".
pref	If not NULL, a character vector of prefixes which may result in erroneously low JW distances. Synonyms will only be considered if both terms share the same prefix
suff	If not NULL, a character vector of suffices which may result in erroneously low JW distances. Synonyms will only be considered if both terms share the same suffix
exclude	If not NULL, a character vector of group names which should be skipped - useful for groups which are known to contain potentially similar terms
verbose	A logical determining if function progress be reported using the pbapply progress bar

### Value

a dataframe of synonyms (cols 1 and 2), the group in which they occur, the frequencies of each synonym in the dataset and finally the q-gram difference between the synonyms

### Examples

```
# load dataset
data("brachios")
# define suffixes
b_suff <- c("ina", "ella", "etta")
# run function
spl <- spell_check(brachios, terms = "genus", groups = "family", suff = b_suff)
```

---

tgraph	<i>tgraph</i>
--------	---------------

---

### Description

Function to create a tgraph representation of a hierarchically organised dataframe. This is the focal object of the t\* functions - the complete set of hierarchical relationships between a set of elements

### Usage

```
tgraph(x, ranks = NULL, verbose = TRUE)
```

**Arguments**

x	A dataframe containing a set of hierarchical relationships. The leftmost column contains the elements which will form the highest rank, followed rightwards by successive ranks
ranks	If not NULL, a vector of column names of x, given in rank order. This is useful if x contains columns which are not rank relevant or if columns are not in hierarchical order. If not supplied, the column order in x is used directly and is assumed to be in rank order
verbose	A logical indicating whether the progress of tgraph construction should be reported to the console

**Value**

a tgraph object

---

threshold_peaks	<i>threshold_peaks</i>
-----------------	------------------------

---

**Description**

Function to detect if two peaks in a density spectrum can be considered separate based on a user supplied threshold. Creates a sequence of divisions from the troughs immediately preceding any significant peaks, then bins occurrences for a given taxon name by those divisions.

**Usage**

```
threshold_peaks(
  x,
  y,
  ycols = c("genus", "max_ma", "min_ma"),
  thresh = 15,
  verbose = TRUE
)
```

**Arguments**

x	A list of significant peaks as returned by find_peaks
y	An occurrence dataset with taxon names corresponding to the list names of x
ycols	A character vector denoting, in order, the taxon, FAD and LAD columns in y
thresh	The threshold distance between peaks above which they will be considered distinct - given in Ma
verbose	A logical determining if function progress should be reported



---

threshold_ranges	<i>threshold_ranges</i>
------------------	-------------------------

---

## Description

Function to detect if two peaks in a density spectrum can be considered separate based on a user supplied threshold. Creates a sequence of divisions from the troughs immediately preceding any significant peaks, then bins occurrences for a given taxon name by those divisions.

## Usage

```
threshold_ranges(
  x,
  rank = "genus",
  srt = "max_ma",
  end = "min_ma",
  method = "kernel",
  step = 1,
  density = 0.1,
  use_sd = TRUE,
  win = 5,
  thresh = 5,
  ...,
  report = TRUE,
  verbose = TRUE
)
```

## Arguments

x	An occurrence dataset containing taxon names, maximum ages and minimum ages
rank	The column name in x containing the taxon names
srt	A column name in x denoting the occurrence maximum ages
end	A column name in x denoting the occurrence minimum ages
method	The method for quantifying occurrence density: one histogram or kernel. Kernel is the recommended default. As called by @seealso densify
step	A positive integer specifying the time window size for density calculation. As called by @seealso densify
density	A positive numeric specifying the step size for densifying records. This should ideally be smaller than step. As called by @seealso densify
use_sd	A logical determining whether to use peaks detected as significant using the mean + standard deviation of its neighbourhood. If FALSE, then the peaks need only be greater than the neighbourhood mean to be significant. Thus, use_sd is more conservative, but less prone to noise. As called by @seealso find_peaks

win	A positive integer specifying the neighborhood window length on either side of a peak during significance testing (i.e. win 5 will give a total window of 11: -5 indices + peak index + 5 indices). As called by @seealso find_peaks
thresh	The threshold distance between peaks above which they will be considered distinct - given in Ma
...	additional arguments passed to @seealso density
report	A logical determining if the analytical outputs of the function be returned to the user, as well as the revised taxon names, TRUE by default
verbose	A logical determining if function progress should be reported

### Value

If report = TRUE (the default), a list of five elements. \$data gives the thresholded (and potentially subdivided) taxon names. \$matrix is the taxon-wise matrix of occurrence densities. \$peaks is a list containing three lists of peaks (all peaks, significant by mean + sd, significant by sd only) for each taxon and a dataframe of peak counts between the three treatments. \$comparison

### Examples

```
# load dataset
data("brachios")
# subsample brachios to make for a short example runtime
set.seed(1)
brachios <- brachios[sample(1:nrow(brachios), 1000),]
# interpeak thresholding
itp <- threshold_ranges(brachios, win = 8, thresh = 10,
                        rank = "genus", srt = "max_ma", end = "min_ma")
```

---

update\_graph

*update\_graph*

---

### Description

Function to update the structure of a graph, given a set of modification as returned by assess\_duplicates

### Usage

```
update_graph(x, del = NULL, add = NULL, changes = NULL)
```

### Arguments

x	a tgraph object to modify
del	A vector of element names or numbers to delete
add	An edge sequence of edges to add to the graph
changes	Alternatively, the output of assess_duplicates, containing proposed deletions and additions

*update\_graph*

35

**Value**

An updated tgraph object

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